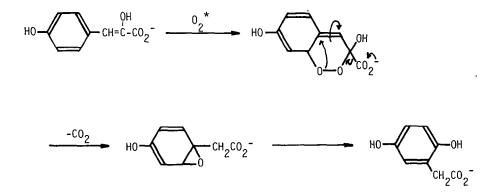
PHOTOSENSITIZED OXYGENATION OF PHENYLPYRUVIC ACID DERIVATIVES AS A MODEL FOR *p*-Hydroxyphenylpyruvate dioxygenase¹

Hıyoshizo Kotsukı, Isao Saito,* and Teruo Matsuura Department of Synthetic Chemistry, Faculty of Engineering, Kyoto University Kyoto 606, Japan

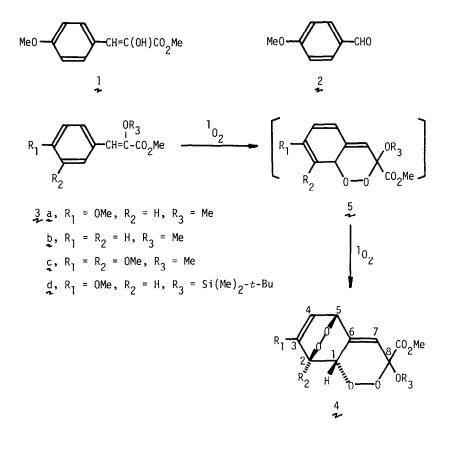
<u>Summary</u>: Dye-sensitized photooxygenation of enol ethers of methyl phenylpyruvates (3) produced diendoperoxides 4 in aprotic solvents, whereas in methanol hydroxylated product 6 was obtained. The reaction scheme is discussed in comparison with that for the enzymic transformation.

The mechanism of the conversion of p-hydroxyphenylpyruvate into homogentisate catalyzed by dioxygenase has been a subject of current interest.^{2,3} At least three different reaction sequences have been proposed for the mechanism of the enzymic reaction on the basis of related chemical model reactions.^{2,3} We previously proposed a mechanism involving 1,4-addition of molecular oxygen to the enol tautomer of p-hydroxyphenylpyruvate as exemplified in Scheme 1.^{3b} In order to obtain a chemical support for the proposed mechanism, we have investigated the reaction of the enol ethers with singlet oxygen.

Scheme 1

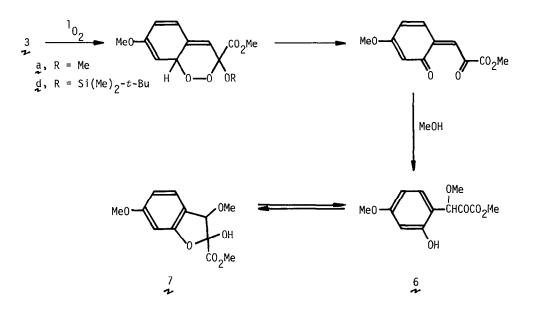


Methylene blue- or rose bengal-sensitized photooxygenation of methyl *p*-methoxyphenylpyruvate (1) in methanol proceeded sluggishly with the bleaching of dye to give *p*-methoxybenzaldehyde (2) and monomethyl oxalate as reported previously.⁴ However, photooxygenation of enol ether derivatives 3 resulted in smooth formation of diendoperoxides 4. For example, photooxygenation of 3a (7.5 mM) in CCl₄ in the presence of tetraphenylporphyrin (TPP) at 0 °C followed by rapid preparative TLC (silica gel) provided an unstable endoperoxide 4a (15%) together with a minor amount of 2 (6%). The structure of 4a was assigned on the basis of spectral data including ¹H and ¹³C NMR.⁵ The chemical shifts and coupling constants of the proton signals are in good agreement with those of the model compounds reported by Matsumoto *et al.*⁶ TPP-sensitized photooxygenation of 3b and 3c in CCl₄ under similar conditions gave 4b⁷ (20%) and 4c⁷ (15%), respectively. Attempts to obtain monoendoperoxide 5 at low conversion or at low-temperature photooxygenation have been unsuccessful. An analogous type of 1,4-cycloaddition of singlet oxygen to vinyl-substituted aromatics has been extensively studied by Foote and Matsumoto *et al.*⁸



The photooxygenation in methanol, however, produced a different type of product. Methylene blue-sensitized photooxygenation of $\frac{3}{4}$ (7 mM) in methanol at 0 °C afforded $\frac{6}{7}$ (50% based on consumed $\frac{3}{4}$): none of the endoperoxides such as $\frac{4}{4}$ has been detected. The product 6 exists in an equilibrated mixture of $\frac{6}{6}$ and $\frac{7}{7}$ in d₆-acetone as evidenced by ¹H NMR. Likewise, photo-oxygenation of $\frac{tert}{t}$ -butyldimethylsilyl ether $\frac{3}{4}$ in methanol afforded $\frac{6}{6}$ (20%) and $\frac{2}{2}$ (20%). The formation of $\frac{6}{6}$ may be rationalized by the mechanism shown in Scheme 2 as proposed previously.⁹

Scheme 2



The present work described here indicates that the enol tautomer of p-hydroxyphenylpyruvate is capable of undergoing 1,4-addition with singlet oxygen under proper conditions when the enol is protected. However, the reaction sequence of Scheme 2 is quite different from that for the enzymic transformation, *i.e.*, the foregoing chemical oxygenation results in *metha*hydroxylation, whereas the enzymic reaction induces *para*-hydroxylation with the migration of the alkyl side chain. Nevertheless, the present result does not necessarily rule out the mechamism of Scheme 1 for the enzymic reaction.

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- 5. Mp 79 80 °C; ^IH NMR (CDC1₃) δ 3.46 (s, 3 H), 3.61 (s, 3 H), 3.78 (s, 3 H), 4.72 (m, 1 H, C-2H), 5.24 (dd, 1 H, J = 3.3, 2.0 Hz, C-1H), 5.31 (1 H, d, J = 8 Hz, C-5H), 5.51 (dd, 1 H, J = 8 Hz, 2.5 Hz, C-4H), 5.68 (d, 1 H, J = 2 Hz, C-7H); ¹³C NMR (CDC1₃) δ 52.6 (q), 52.8 (q), 55.4 (q), 72.7 (d), 73.7 (d), 73.9 (d), 96.8 (d), 101.2 (s), 110.6 (d), 145.5 (s), 156.6 (s), 167.1 (s).
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- 7. All new compounds exhibited consistent spectral data $({}^{1}H \text{ and } {}^{13}C \text{ NMR, MS})$ and elemental analyses. Selected spectral data follow. 4b: mp 123 - 124.5 °C; ${}^{1}H \text{ NMR} (\text{CDCl}_3) \delta 3.41$ (s, 3 H), 3.71 (s, 3 H), 4.76 (m, 1 H, C-2H), 5.08 (dt, 1 H, J = 6.5, 1.2 Hz, C-5H), 5.17 (dd, 1 H, J = 3, 2 Hz, C-1H), 5.71 (d, 1 H, J = 2 Hz, C-7H), 6.40 (ddd, 1 H, J = 8.3, 5.8, 1.2 Hz, C-3H), 6.79 (ddd, 1 H, J = 8.3, 6.5, 1.7 Hz, C-4H). 4c: mp 120 - 121.5 °C; ${}^{1}H \text{ NMR} (\text{CDCl}_3)$ $\delta 3.48$ (s, 3 H), 3.66 (s, 3 H), 3.77 (s, 3 H), 3.78 (s, 3 H), 5.26 (d, 1 H, J = 7 Hz, C-5H), 5.38 (d, 1 H, J = 2 Hz, C-1H), 5.53 (d, 1 H, J = 7 Hz, C-4H), 5.71 (d, 1 H, J = 2 Hz, C-7H). 6: mp 112 - 113 °C; ${}^{1}H \text{ NMR} (\text{CDCl}_3) \delta 3.57$ (s, 3 H), 3.76 (s, 3 H), 3.84 (s, 3 H), 5.01 (s, 1 H), 5.18 (br, 1 H, OH), 6.44 (d, 1 H, J = 2.5 Hz), 6.53 (dd, 1 H, J = 8, 2.5 Hz), 7.24 (d, 1 H, J = 8 Hz).
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